The content of the paper and computer readable copies of the Sequence Listing submitted in accordance with 37 C.F.R. §1.821(c) and (e) are the same and do not include new matter.

The Office Communication contains an objection to the use of the variable nucleotide "n" in SEQ ID NO:12 absent entries of features <220> to <223>. Applicants discovered that the sequence provided as SEQ ID NO: 12 was recited in error in the Sequence Listing provided on June 15, 2001. The basis for this error, the sequence recited as associated with human tumor necrosis factor ("Hu-TNF") in Figure 2C, lacks the sequence "sunc" and therefore lacks the variable nucleotide. The sequence in Figure 2C was correctly recited in the Sequence Listing submitted March 30, 2001. However, for reasons that are unknown (this application being transferred to our law firm in July-August 2001), the Sequence Listing submitted on June 15, 2001 erroneously recited the Hu-TNF sequence plus "sunc" at the 5' end, a clear error. Applicants have now corrected the error in SEQ ID NO:12, and the correction, because it restores the sequence to its original state, does not constitute new matter. Furthermore, the basis for the present Office Communication is obviated, because there is no variable nucleotide.

In addition, it was discovered that the Sequence Listings submitted June 15, 2001 and March 30, 2001 contained errors in SEQ ID NO:8, wherein, instead of the 20 bp oligonucleotide sequence provided in the specification as originally filed, recited a 22 bp sequence. This has now been corrected so that the Sequence Listing conforms to the specification. As such, the correction does not constitute new matter.

A replacement paragraph appears in the preceding "IN THE SPECIFICATION" section. Attached hereto is a marked-up version of the changes made

to the specification paragraph, on a page captioned "<u>VERSION WITH MARKINGS TO SHOW CHANGES MADE</u>." Should any discrepancies be discovered, the version presented in the preceding "IN THE SPECIFICATION" section shall take precedence.

As attorneys for Applicant have changed, this paper is submitted and signed pursuant to 37 C.F.R. §1.34(a). An executed Revocation and Power of Attorney will follow.

Please charge any fees additional fees associated with this submission or credit any overpayment to Deposit Account No. 02-4377. Two copies of this paper are enclosed.

Respectfully submitte

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IN THE SPECIFICATION

In the Experimental Details, the paragraph on page 59, line 20 through page 60, line 12 has been amended as follows:

Electrophoretic Mobility Shift Assay (EMSA). Nuclear extracts were prepared as described (25). Briefly, binding reactions were performed in 10 or 20 ml reaction mixtures containing 1-3 mg of nuclear extracts from control or differentiation inducer treated HO-1 cells. The binding buffer contained 12 mM HEPES (pH 7.9), 5 mM MgCl₂, 60 mM KCl, 0.6 mM EDTA, 0.5 mM dithiothreitol, 1 mg of poly (dI-dC), 10% glycerol. The region corresponding to the putative AP-1 and C/EBP binding sites present between NdeI and NheI restriction enzyme sites was PCR amplified using flanking primers, 5'-AGGCTGGATTTG GCTTGTGAC-3' (Sense) (SEQ NO:3) IDCTGTTTAATCCAGCACTTCCC-3' (Antisense) (SEQ ID NO:4). The PCR product was column purified (Oiagen), end labeled with α -32P [ATP] and 1500 cpm of double stranded DNA were used per binding reaction. Binding reactions were performed at RT for 30 min. Reactions were then loaded onto a 4% polyacrylamide gel and electrophoresed at 4° C at 100 V in 0.25X Tris-borate-EDTA as described (26, 27). Competition and supershift reactions were identical to those described above, except a 10-100 fold excess of AP-1 or [C/EPB] C/EBP wild type or mutant oligonucleotides (AP-1/WT; 5'-CGCTTGATGACTCAGCCGGAA-3'), (SEQ ID NO: 5), (C/EBP/WT; 5'-TGCAGATTGCGCAATCTGC A-3'), (SEQ ID NO. 6), (AP-1/MT;CGCTTGATGACTTGGCCGGAA-3') (SEQ ID NO:7) and C/EBP (C/EBP/MT; 5'-TGCAGAGACTAGTCTCTGCA-3') (SEQ ID NO:8) or 1-5 µg of either anti-cJun/AP-1

or [anti-C/EPB] anti-C/EBP antibody (Santa Cruz) were added to the binding reactions along with the labeled probe and reactions were incubated for 30 min at RT prior to electrophoresis. The gels were then dried and exposed to X-ray film.